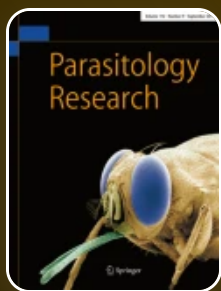


[Home](#) > [Parasitology Research](#) > [Article](#)

Blood parasites of passerines in the Brazilian Pampas and their implications for a potential population supplementation program for the endangered Yellow Cardinal (*Gubernatrix cristata*)

| Research | Published: 03 September 2022


| Volume 121, pages 3203–3215, (2022) [Cite this article](#)



[Parasitology Research](#)

[Aims and scope](#) →

[Submit manuscript](#) →

Bianca Ressetti da Silva, Ralph Eric Thijl Vanstreels, Patricia P. Serafini, Carla Suertegaray Fontana, Thaiane Weinert da Silva, Eduardo Chiarani, Andréa M. Carvalho, Francisco C. Ferreira Junior, Érika Martins Braga & Rosangela Locatelli-Dittrich 

 487 Accesses  1 Citation  2 Altmetric

[Explore all metrics](#) →

Abstract

Espinilho savanna (“seasonal steppe savanna”) is a unique vegetation formation of the Pampas biome that is found near the tri-border of Brazil, Uruguay, and Argentina. The Yellow Cardinal (*Gubernatrix cristata*) is a flagship species of this ecosystem, but it is classified as “critically endangered” in Brazil due to habitat loss and poaching for the illegal trade. Population supplementation through the release of individuals that were captive-bred or apprehended by authorities from the illegal trade has been considered as a conservation strategy for this species; however, the risk of

pathogen introduction is a critical concern. We used microscopy and molecular methods to investigate the occurrence of blood parasites in wild passerines ($n = 64$, including three Yellow Cardinals) at Espinilho State Park, Rio Grande do Sul, Brazil, and in captive Yellow Cardinals ($n = 30$) at three facilities in Brazil. Haemosporidian parasites were detected in the blood smears of 10.9% of the wild passerines, comprising the morphospecies *Haemoproteus erythrogravidus* in Rufous-collared Sparrow (*Zonotrichia capensis*), *H. quiscalus* in Grayish Baywing (*Agelaioides badius*), and *H. tyranni* in Great Kiskadee (*Pitangus sulphuratus*); these are the southernmost records for these morphospecies and their first record for the Pampas biome. No haemosporidian parasites were detected in the blood smears of the Yellow Cardinals, wild or captive. Microfilariae were detected in the blood smears of 14.1% of the wild passerines, including all wild Yellow Cardinals, and in 43.3% of captive Yellow Cardinals. *Trypanosoma* sp. was detected in the blood smear of one captive Yellow Cardinal. Nested PCR and gene sequencing of the *cyt-b* gene of *Haemoproteus/Plasmodium* was used to test a subset of wild passerines and captive Yellow Cardinals, allowing for the

molecular barcoding of *H. quiscalus* lineage AGEBAD04 and *H. tyranni* lineage PITSUL01; additionally, DNA identical to that of lineage PITSUL01 was detected in the blood of one captive Yellow Cardinal. This study provides valuable data to support the conservation management of the Yellow Cardinal and other threatened passerines from the Pampas and highlights the need for further studies on the epidemiology and pathology of filarioid worms and trypanosomes in passerines from this biome.

 This is a preview of subscription content, [log in via an institution](#)  to check access.

Access this article

[Log in via an institution](#) →

[Buy article PDF 39,95 €](#)

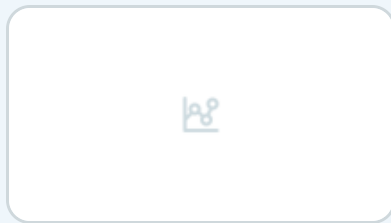
Price includes VAT (Bangladesh)

Instant access to the full article PDF.

Rent this article via [DeepDyve](#) ↗

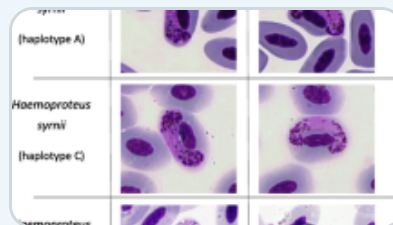
[Institutional subscriptions](#) →

Similar content being viewed by others



Low prevalence of haemosporidian parasites in...

Article | 28 June 2018



Haemosporidian parasites from captive Strigiformes i...

Article | 18 July 2020



A molecular survey of vector-borne...

Article | Open access
08 February 2015

Data availability

Genetic sequences produced in this study were deposited at GenBank (accession codes ON703102–4) and MalAvi (lineages AGEBAD04 and PITSUL01).

References

Atawal AF, Mgbeahuruike AC, Hammers M (2019) Microfilarial infections associated with body mass loss of Village Weavers *Ploceus cucullatus*. *Ostrich* 90:41–44.
<https://doi.org/10.2989/00306525.2018.1539418>

[Article](#) [Google Scholar](#)

Beier C, Repenning M, da Pereira MS et al (2017) Cooperative breeding and demography of Yellow Cardinal *Gubernatrix cristata* in Brazil. *Rev Bras Ornitol* 25:12–19.
<https://doi.org/10.1007/BF03544371>

[Article](#) [Google Scholar](#)

Bennett GF, Campbell A (1972) Avian Haemoproteidae. I. Description of *Haemoproteus fallisi* n. sp. and a review of the

haemoproteids of the family Turdidae. Can J Zool 50:1269–1275

[Article](#) [CAS](#) [Google Scholar](#)

Bennett GF, Caines JR, Whiteway MA (1986) Avian Haemoproteidae. 20. The haemoproteids of the avian families Apodidae (swifts), Bucconidae (puffbirds), and Indicatoridae (honeyguides). Can J Zool 64:766–770.

<https://doi.org/10.1139/z86-113>

[Article](#) [Google Scholar](#)

Bensch S, Hellgren O, Pérez-Tris J (2009) MalAvi: a public database of malaria parasites and related haemosporidians in avian hosts based on mitochondrial cytochrome b lineages. Mol Ecol Resour 9:1353–1358

[Article](#) [Google Scholar](#)

BirdLife International (2022) Species factsheet:

Gubernatrix cristata. In: BirdLife International Data Zone.

<http://datazone.birdlife.org/species/factsheet/yellow-cardinal-gubernatrix-cristata>. Accessed 6 Jun 2022

Burphy-Caines JR, Bennett GF (1992) The Haemoproteidae (Apicomplexa: Haemosporina) of the avian families Fringillidae and Emberizidae s.l. Can J Zool 70:1149–1160. <https://doi.org/10.1139/z92-161>

[Article](#) [Google Scholar](#)

Clark NJ, Adlard RD, Clegg SM (2015) Molecular and morphological characterization of *Haemoproteus* (*Parahaemoproteus*) *ptilotis*, a parasite infecting Australian honeyeaters (Meliphagidae), with remarks on prevalence and potential cryptic speciation. Parasitol Res 114:1921–1928

[Article](#) [Google Scholar](#)

Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: more models, new heuristics and parallel

computing. Nat Methods 9:772–772.

<https://doi.org/10.1038/nmeth.2109>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

de Castaneda RR, Morales J, Moreno Klemming J et al (2009) Costs and benefits of early reproduction: *Haemoprotus* prevalence and reproductive success of infected male pied flycatchers in montane habitat in Central Spain. Ardeola 56:271–280

[Google Scholar](#)

de Oliveira L, Barino GTM, Rossi MF et al (2020) Morphological and molecular characterization of *Haemoproteus coatneyi* and *Haemoproteus erythrogravidus* (Haemosporida: Haemoproteidae) in Passeriformes in Brazil's Atlantic Forest. Rev Bras Parasitol Veterinária 29:e011520. <https://doi.org/10.1590/s1984-29612020074>

[Article](#) [CAS](#) [Google Scholar](#)

De La Torre GM, Campiã KM (2021) Bird habitat preferences drive hemoparasite infection in the Neotropical region. *Integr Zool* 16:755–768.

<https://doi.org/10.1111/1749-4877.12515>

[Article](#) [CAS](#) [Google Scholar](#)

Dimitrov D, Valkiūnas G, Zehtindjiev P et al (2013) Molecular characterization of haemosporidian parasites (Haemosporida) in yellow wagtail (*Motacilla flava*), with description of in vitro ookinetes of *Haemoproteus motacillae*. *Zootaxa* 3666:369–381

[Article](#) [Google Scholar](#)

Doussang D, González-Acuña D, Torres-Fuentes LG et al (2019) Spatial distribution, prevalence and diversity of haemosporidians in the rufous-collared sparrow *Zonotrichia Capensis*. *Parasit Vectors* 12:2.

<https://doi.org/10.1186/s13071-018-3243-4>

[Article](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

Doussang D, Sallaberry-Pincheira N, Cabanne GS et al (2021) Specialist versus generalist parasites: the interactions between host diversity, environment and geographic barriers in avian malaria. *Int J Parasitol* 51:899–911. <https://doi.org/10.1016/j.ijpara.2021.04.003>

[Article](#) [PubMed](#) [Google Scholar](#)

Durrant KL, Beadell JS, Ishtiaq F et al (2006) Avian hematozoa in South America: a comparison of temperate and tropical zones. *Ornithol Monogr* 60:98–111. <https://doi.org/10.2307/40166831>

[Article](#) [Google Scholar](#)

Ellis VA, Fecchio A, Ricklefs RE (2020) Haemosporidian parasites of neotropical birds: causes and consequences of infection. *Auk* 137:ukaa055. <https://doi.org/10.1093/auk/ukaa055>

[Article](#) [Google Scholar](#)

Fecchio A, Marini MÂ, Braga ÉM (2007) Baixa prevalência de hemoparasitos em aves silvestres no Cerrado do Brasil Central. *Neotropical Biol Conserv* 2:127–135

[Google Scholar](#)

Fecchio A, Bell JA, Pinheiro RBP et al (2019) Avian host composition, local speciation and dispersal drive the regional assembly of avian malaria parasites in South American birds. *Mol Ecol* 28:2681–2693.

<https://doi.org/10.1111/mec.15094>

[Article](#) [PubMed](#) [Google Scholar](#)

González AD, Matta NE, Ellis VA et al (2014) Mixed species flock, nest height, and elevation partially explain avian haemoparasite prevalence in Colombia. *PLoS ONE* 9:e100695. <https://doi.org/10.1371/journal.pone.0100695>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

Hamilton PB, Gibson WC, Stevens JR (2007) Patterns of co-evolution between trypanosomes and their hosts deduced from ribosomal RNA and protein-coding gene phylogenies. *Mol Phylogenet Evol* 44:15–25.

<https://doi.org/10.1016/j.ympev.2007.03.023>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

Hellgren O, Waldenström J, Bensch S (2004) A new PCR assay for simultaneous studies of *Leucocytozoon*, *Plasmodium*, and *Haemoproteus* from avian blood. *J Parasitol* 90:797–802

[Article](#) [CAS](#) [Google Scholar](#)

Huang Y-L, Tsai S-S, Thongchan D et al (2017) Filarial nematode infection in eclectus parrots (*Eclectus roratus*) in Taiwan. *Avian Pathol* 46:188–194.

<https://doi.org/10.1080/03079457.2016.1237014>

Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio) (2018) Livro Vermelho da Fauna Brasileira Ameaçada de Extinção: Volume III – Aves. Ministério do Meio Ambiente, Brasília, Brazil.

https://www.gov.br/icmbio/pt-br/centrais-de-conteudo/publicacoes/publicacoes-diversas/livro_vermelho_2018_vol3.pdf

Iezhova TA, Valkiūnas G, Loiseau C et al (2010) *Haemoproteus cyanomitrae* sp. nov. (Haemosporida: Haemoproteidae) from a widespread African songbird, the olive sunbird *Cyanomitra Olivacea*. J Parasitol 96:137–143

Jaramillo A (2020) Yellow Cardinal (*Gubernatrix cristata*).
In: del Hoyo J, Elliott A, Sargatal J, de Juana E (eds) Birds of
the world. Cornell Lab of Ornithology, Ithaca, NY.

<https://doi.org/10.2173/bow.yelcar1.01>

Križanauskienė A, Pérez-Tris J, Palinauskas V et al (2010)
Molecular phylogenetic and morphological analysis of
haemosporidian parasites (Haemosporida) in a naturally
infected European songbird, the blackcap *Sylvia atricapilla*,
with description of *Haemoproteus pallidulus* sp. nov.

Parasitology 137:217–227

[Article](#) [Google Scholar](#)

Križanauskienė A, Iezhova TA, Palinauskas V et al (2012)
Haemoproteus nucleocondensus n. sp. (Haemosporida,
Haemoproteidae) from a Eurasian songbird, the Great Reed
Warbler *Acrocephalus arundinaceus*. Zootaxa 3441:36–46

[Article](#) [Google Scholar](#)

Kumar S, Stecher G, Li M et al (2018) MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol* 35:1547–1549.

<https://doi.org/10.1093/molbev/msy096>

[Article](#) [CAS](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

Laird M, Van Riper C (1981) Questionable reports of *Plasmodium* from birds in Hawaii, with the recognition of *P. relictum* ssp. *capistranoae* (Russell, 1932) as the avian malaria parasite there. In: Canning EW (ed) Parasitological topics - a presentation volume to P.C.C. Garnham, F.R.S. on the occasion of his 80th birthday. Society of Protozoologists. Lawrence, Kansas, pp 159–165

Law JM, Tully TN, Stewart TB (1993) Verminous encephalitis apparently caused by the filarioid nematode *Chandlerella quiscali* in emus (*Dromaius novaehollandiae*). *Avian Dis* 37:597–601

[Article](#) [CAS](#) [Google Scholar](#)

Mantilla JS, González AD, Lotta IA et al (2016)
Haemoproteus erythrogravidus n. sp. (Haemosporida,
Haemoproteidae): description and molecular
characterization of a widespread blood parasite of birds in
South America. *Acta Trop* 159:83–94

[Article](#) [Google Scholar](#)

Marchiori JNC, Alves FDS (2014) O inhanduvá (*Prosopis
affinis* Spreng.) no Rio Grande do Sul. 8 – Aspectos
fitogeográficos. *Balduinia* 0:13–20.

<https://doi.org/10.5902/2358198014133>

[Article](#) [Google Scholar](#)

Martinsen ES, Perkins SL, Schall JJ (2008) A three-genome
phylogeny of malaria parasites (*Plasmodium* and closely
related genera): evolution of life-history traits and host
switches. *Mol Phylogenet Evol* 47:261–273

[Article](#) [CAS](#) [Google Scholar](#)

Martins-Ferreira C, Freitas TRO (2010) Genetic diversity and population structure of the endangered yellow cardinal *Gubernatrix cristata* and implications to its conservation efforts. In: Proceedings of the 25th International Ornithological Congress. International Ornithological Congress. Campos do Jordão, Brazil, p 868

Marzal A, Ricklefs RE, Valkiūnas G et al (2011) Diversity, loss, and gain of malaria parasites in a globally invasive bird. PLoS ONE 6:e21905

[Article](#) [CAS](#) [Google Scholar](#)

Nandi N, Bennett GF (1994) Re-description of *Trypanosoma corvi* Stephens and Christophers, 1908 emend. Baker, 1976 and remarks on the trypanosomes of the avian family Corvidae. Mem Inst Oswaldo Cruz 89:145–151

[Article](#) [Google Scholar](#)

Olson DM, Dinerstein E, Wikramanayake ED et al (2001) Terrestrial ecoregions of the world: a new map of life on Earth. *Bioscience* 51:933. [https://doi.org/10.1641/0006-3568\(2001\)051\[0933:TEOTWA\]2.0.CO;2](https://doi.org/10.1641/0006-3568(2001)051[0933:TEOTWA]2.0.CO;2)

[Article](#) [Google Scholar](#)

Palinauskas V, Žiegytė R, Iezhova TA et al (2016) Description, molecular characterisation, diagnostics and life cycle of *Plasmodium elongatum* (lineage pERIRUB01), the virulent avian malaria parasite. *Int J Parasitol* 46:697–707. <https://doi.org/10.1016/j.ijpara.2016.05.005>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

Paperna I, Yosef R (2010) Description of a new species of *Haemoproteus* (Haemosporidia, Apicomplexa) from an Orange-tufted Sunbird *Nectarinia osea*. *Acta Parasitol* 55:103–107

[Article](#) [Google Scholar](#)

Piacentini VQ, Aleixo A, Agne CE et al (2015) Annotated checklist of the birds of Brazil by the Brazilian Ornithological Records Committee/Lista comentada das aves do Brasil pelo Comitê Brasileiro de Registros Ornitológicos. Rev Bras Ornitol 23:91–298

[Article](#) [Google Scholar](#)

Redin CG, Longhi RV, Watzlawick LF, Longhi SJ (2011) Composição florística e estrutura da regeneração natural do Parque Estadual do Espinilho, RS. Ciênc Rural 41:1195–1201

[Article](#) [Google Scholar](#)

Ribeiro SF, Sebaio F, Branquinho FCS et al (2005) Avian malaria in Brazilian passerine birds: parasitism detected by nested PCR using DNA from stained blood smears. Parasitology 130:261–267.

<https://doi.org/10.1017/S0031182004006596>

[Article](#) [CAS](#) [PubMed](#) [Google Scholar](#)

Ronquist F, Teslenko M, Van Der Mark P et al (2012)
MrBayes 3.2: efficient Bayesian phylogenetic inference and
model choice across a large model space. Syst Biol 61:539–
542. <https://doi.org/10.1093/sysbio/sys029>

[Article](#) [PubMed](#) [PubMed Central](#) [Google Scholar](#)

Sambrook J, Russell D (2001) Molecular cloning: a
laboratory manual, 3rd edn. Cold Spring Harbor Laboratory
Press, Cold Spring Harbor

[Google Scholar](#)

Schneider CA, Rasband WS, Eliceiri KW (2012) NIH Image
to ImageJ: 25 years of image analysis. Nat Methods 9:671–
675

[Article](#) [CAS](#) [Google Scholar](#)

Sebaio F, Braga ÉM, Branquinho F et al (2012) Blood
parasites in passerine birds from the Brazilian Atlantic

Forest. Rev Bras Parasitol Veterinária 21:7–15.

<https://doi.org/10.1590/S1984-29612012000100003>

[Article](#) [Google Scholar](#)

Serafini PP (2013) Plano de Ação Nacional para a Conservação dos Passeriformes Ameaçados dos Campos Sulinos e Espinilho. Instituto Chico Mendes de Conservação da Biodiversidade, Brasília, Brazil.

https://www.icmbio.gov.br/cemave/images/stories/Publicações_científicas/livro-campos-sulinos-2013.pdf

Tostes R, Vashist U, Scopel KKG et al (2015) *Plasmodium* spp. and *Haemoproteus* spp. infection in birds of the Brazilian Atlantic Forest detected by microscopy and polymerase chain reaction. Pesqui Veterinária Bras 35:67–74. <https://doi.org/10.1590/S0100-736X2015000100014>

[Article](#) [Google Scholar](#)

Valkiūnas G (2005) Avian malaria parasites and other haemosporidia. CRC Press, Boca Raton

[Google Scholar](#)

Valkiūnas G, Žičkus T, Shapoval AP, Lezhova TA (2006) Effect of *Haemoproteus belopolskyi* (Haemosporida: Haemoproteidae) on body mass of the blackcap *Sylvia atricapilla*. J Parasitol 92:1123–1125.

<https://doi.org/10.1645/GE-3564-RN.1>

[Article](#) [PubMed](#) [Google Scholar](#)

Valkiūnas G, Križanauskienė A, Iezhova TA et al (2007) Molecular phylogenetic analysis of circumnuclear hemoproteids (Haemosporida: Haemoproteidae) of sylviid birds, with a description of *Haemoproteus parabelopolskyi* sp. nov. J Parasitol 93:680–687.

<https://doi.org/10.1645/GE-1102R.1>

[Article](#) [PubMed](#) [Google Scholar](#)

Viggers K, Lindenmayer D, Spratt D (1993) The importance of disease in reintroduction programmes. *Wildl Res* 20:687.
<https://doi.org/10.1071/WR9930687>

[Article](#) [Google Scholar](#)

White EM, Greiner EC, Bennett GF, Herman CM (1978) Distribution of the hematozoa of Neotropical birds. *Rev Biol Trop* 26:43–102

[PubMed](#) [Google Scholar](#)

Woodworth-Lynas CB, Caines JR, Bennett GF (1989) Prevalence of avian haematozoa in São Paulo state, Brazil. *Mem Inst Oswaldo Cruz* 84:515–526.

<https://doi.org/10.1590/S0074-02761989000400009>

[Article](#) [Google Scholar](#)

Zídková L, Cepicka I, Szabová J, Svobodová M (2012)
Biodiversity of avian trypanosomes. *Infect Genet Evol*
12:102–112. <https://doi.org/10.1016/j.meegid.2011.10.022>

[Article](#) [PubMed](#) [Google Scholar](#)

Acknowledgements

We wish to thank Secretaria Estadual do Meio Ambiente e Infraestrutura do Rio Grande do Sul (SEMA) and Espinilho State Park staff for permitting to research in a protected area and for all the support during fieldwork and Fazenda São Marcos owners for granting access to one of the study sites. We also thank Pai Passo farm Owners (especially Paulo, Pedro Bastos and Maria), Condomínio Agropecuário Ceolin, and Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio) for their huge support during field expeditions.

Funding

This study was supported by the ICMBio, Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq 422053/2016–3), and Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG APQ-00645–18). FCF was supported by the National Science Foundation grant 1717498 as part of the joint NSF–NIH–USDA Ecology and Evolution of Infectious Diseases program. CSF was supported by CNPq grant 309438/2016–0 and 310608/2019–8. EMB was granted by Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq 304334/2019–7).

Author information

Authors and Affiliations

**Programa de Pós-Graduação em Ciências Veterinárias,
Setor de Ciências Agrárias, Universidade Federal do
Paraná, Rua dos Funcionários 1540, Curitiba, PR, 80035-
050, Brazil**

Bianca Ressetti da Silva & Rosangela Locatelli-Dittrich

**Instituto de Pesquisa e Reabilitação de Animais Marinhos,
Rodovia BR 262 Km 0, Cariacica, ES, 29140-130, Brazil**

Ralph Eric Thijl Vanstreels

Centro Nacional de Pesquisa e Conservação de Aves Silvestres, Instituto Chico Mendes de Conservação da Biodiversidade, Rodovia Jornalista Maurício Sirotski Sobrinho km 1, Florianópolis, SC, 88053-700, Brazil

Patricia P. Serafini

Laboratory of Biomarkers of Aquatic Contamination and Immunochemistry, Federal University of Santa Catarina, Servidão Caminho Do Porto S/N, Florianópolis, SC, 88034-257, Brazil

Patricia P. Serafini

Laboratório de Ornitologia, Museu de Ciências e Tecnologia, Programa de Pós-Graduação Em Ecologia E Evolução da Biodiversidade, Pontifícia Universidade Católica do Rio Grande do Sul, Avenida Ipiranga 6681, Porto Alegre, RS, 90619-900, Brazil

Carla Suertegaray Fontana, Thaianne Weinert da Silva & Eduardo Chiarani

Laboratório de Patologia Clínica Veterinária e Diagnóstico Molecular, Faculdade de Agronomia e Medicina

Veterinária, Universidade de Brasília, Brasília, DF, 70910-900, Brazil

Andréa M. Carvalho

Center for Conservation Genomics, Smithsonian Conservation Biology Institute, Washington, DC, USA

Center for Vector Biology, Rutgers University, New Brunswick, NJ, USA

Francisco C. Ferreira Junior

Departamento de Parasitologia, Universidade Federal de Minas Gerais, Avenida Presidente Antônio Carlos, 6627, Belo Horizonte, MG, 31270-901, Brazil

Érika Martins Braga

Contributions

BRS, PPS, CSF, TWS, EC, and RLD conceived the project.

BRS, PPS, CSF, TWS, and EC conducted field expeditions and collected and processed data and biological samples. BRS,

RETV, FCF, and EMB examined blood smears and

morphologically identified the parasites. AMC, FCF, RETV, and EMB conducted molecular and phylogenetic analyses.

BRS, RETV, and RLD wrote the first manuscript draft and

prepared the tables and figures. All authors reviewed and edited the manuscript and approved its final version.

Corresponding author

Correspondence to [Rosangela Locatelli-Dittrich](#).

Ethics declarations

Competing interests

The authors declare no competing interests.

Ethics approval

The study was approved by the Animal Use Ethics Committee of the Agricultural Sciences Campus of the Universidade Federal do Paraná (CEUA 038/2018) and was conducted under permits issued by the Brazilian Ministry of Environment (SISBIO 53935–6).

Consent to participate

Not applicable.

Consent for publication

Not applicable.

Conflict of interest

The authors declare no competing interests.

Additional information

Section Editor: Leonhard Schnittger.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Supplementary Information

Below is the link to the electronic supplementary material.

[**Supplementary file1 \(PDF 936 KB\)**](#)

Rights and permissions

Springer Nature or its licensor holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

[Reprints and permissions](#)

About this article

Cite this article

da Silva, B.R., Vanstreels, R.E.T., Serafini, P.P. *et al.* Blood parasites of passerines in the Brazilian Pampas and their implications for a potential population supplementation program for the endangered Yellow Cardinal (*Gubernatrix cristata*). *Parasitol Res* **121**, 3203–3215 (2022).

<https://doi.org/10.1007/s00436-022-07638-w>

Received

27 June 2022

Accepted

23 August 2022

Published

03 September
2022

Issue Date

November 2022

DOI

<https://doi.org/10.1007/s00436-022-07638-w>

Keywords

[Haemosporida](#)

[Neotropics](#)

[Pampas](#)

[Passeriformes](#)

[Rhabditida](#)

[Trypanosomatida](#)

